

THE BIOMIMETIC SYNTHESIS OF TRANS-(1 β -H,5 α -H)-GUAIANOLIDES

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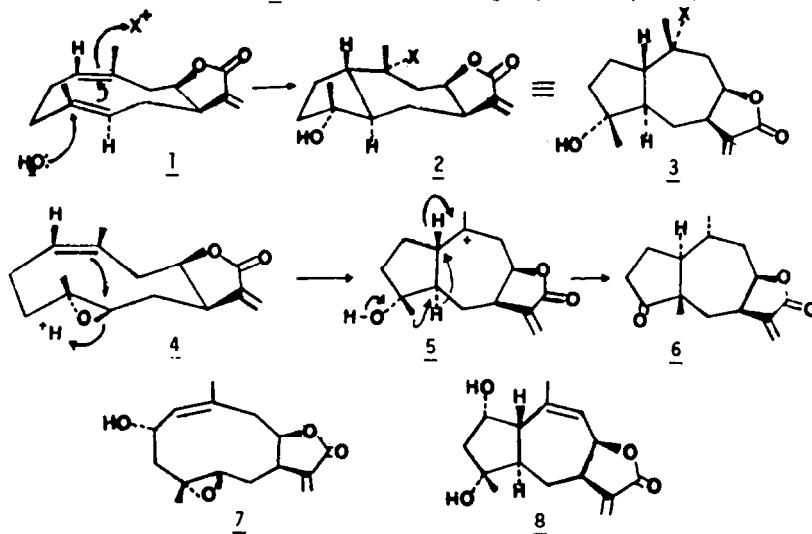
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ABSTRACT: 1-epigallicin 1 underwent biomimetic cyclization to trans-(1 β -H,5 α -H)-guaianolide 22, the stereospecificity of this cyclization being credited to the fact that it took place via the reacting TC conformation 27. The epimeric nor-guaianolides 15 and 17 were prepared and their rearrangements studied. The biogenetic implications of these processes are discussed.

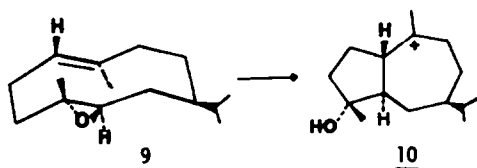
The trans-(1 β -H,5 α -H)-guaianolides 3 are a fairly rare group of sesquiterpene lactones¹,

SCHEME I

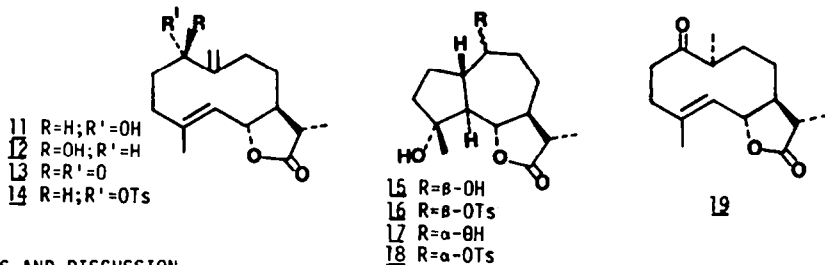


the biogenesis of which has been attributed to an anti-Markownikoff type² trans-antiparallel cyclization via the melampolides 1. Herz³ suggested another possible biogenesis for the trans-guaianolides: 4 α ,5 β -epoxide 4 acid-induced cyclization could give rise to the cation 5, with typical trans-guaianolide stereochemistry and this cation could lead to the helenanolides 6 via the chemical shifts shown in Scheme I. This pathway has recently been favoured since the structural reclassification of baileyin 7 as a melampolide skeleton rather than a germacrolide⁴ together with an X-ray study demonstrating pleniradin 8 to be a trans-guaianolide⁵ strongly suggest that these two lactones isolated from *Baileya pleniradiata* are biogenetically related⁵. Sharma et al⁶ were able to convert a guaianolide into a pseudoguaianolide by imitating part of the proposed biogenetic pathway.

In a review of the melampolides, Fischer et al⁷ suggested that the distance from centre to centre between the double bonds of 1 is considerably greater than in the four possible conformations of the trans,trans-germacradienes and proposed that the trans-guaiane cation 10 must be formed via a 4 α ,5 β -epoxy-trans-germacrene 9 through a quasi-parallel conformation.

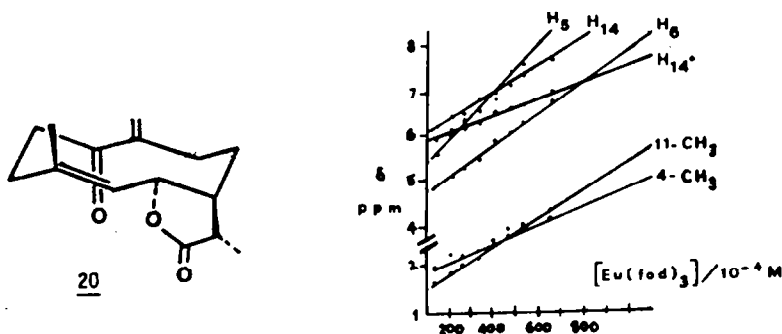


To evaluate the role played by the *cis*,*trans*-germacradienes¹ in the biogenesis of *trans*-guaianolides and the possible implication of **1** in the biogenesis of the helenanolides, we carried out the cyclization of 1-epigallicin **11**⁸ as well as the study of the rearrangements of the nor-*trans*-guaianolides **15** and **17**.



RESULTS AND DISCUSSION

The oxidation of gallicin **12** with activated MnO_2 yielded ketone **13** (77%) which, when reduced with NaBH_4 , gave 1-epigallicin **11**⁹ (57%), the dihydroketone **19** (21%) and gallicin **12** (4%). The stereoselectivity of the reaction could be due to the fact that the reduction took place via the preferred reacting conformation **20**. ¹H-NMR at various temperatures and LIS studies provided the conformational analysis of **13** in solution. The ¹H-NMR spectrum was taken at a standard temperature (35°C) and none of the more important signals changed significantly between -60°C and +60°C. The addition of $\text{Eu}(\text{fod})_3$ caused the chemical shifts shown in Fig. 1. The slight chemical shifts of H-14 in **13** are not compatible with a *syn*-relationship between the ketone group and the double bond 10(14), which suggests an *s-trans* disposition for the α,β -unsaturated ketone. Furthermore, the chemical shift of H-5 suggests a *syn*-axial disposition for H-5 and the ketone group.



In the 2D-NOE study of **13**, positive correlations between H-6/H-15 and H-14/H-15 were observed and clearly indicate the *syn*-disposition of these protons. Furthermore the Cotton effect curve of **13** has a value of $\Delta\epsilon_{\text{max}} = +9.82$ at 240 nm (Band K), which points to a *transoid* arrangement for the chromophore¹⁰

These data agree with the crown conformation **20** in which the "si" face of the ketone group is highly hindered and the hydride ion attack takes place on the "re" face, generating **11**.

Molecular Mechanics calculations made with the program MMP1¹¹ on the CC, CT, TT and TC conformations of **13** give the values 30.17; 31.63; 33.96 and 34.12 kcal/mol clearly showing a conformational preference for the CC conformer (91.93% population).

An X-ray diffraction study of **13** showed this compound to present two crystallographically

independent molecules, identically configured and almost identically conformed if the slight differences in the torsional angles around C(x8)-C(x9) and C(x9)-C(x10) are discounted. A half-normal probability plot¹² calculated from a comparison of all intramolecular distances < 4.0 Å (between non-H atoms and excluding the distances involved in the above-mentioned torsional angles) gives a linear array with a correlation coefficient of 0.99 and zero intercept at -0.06(3). The ten-membered ring has CCC conformation¹³, although slightly distorted owing to the influence of the substituents (Fig. 4). A list of bond lengths, bond angles and torsional angles for compound 13 are given in Table 2. The lactone ring attached to C(x6) and C(x7) shows an envelope conformation, with C(x7) at the flap and 0.562(8) Å (x=1) and 0.444(7) Å (x=2) out of the plane defined by the other four atoms.

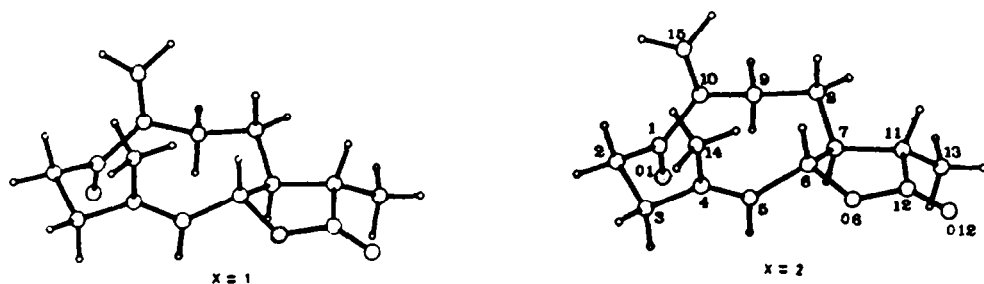


Figure 2. A view of the two crystallographically independent molecules of compound 13. The atomic numbering is arrived at by placing the indicated value of x at the beginning of the numerals shown in the figure.

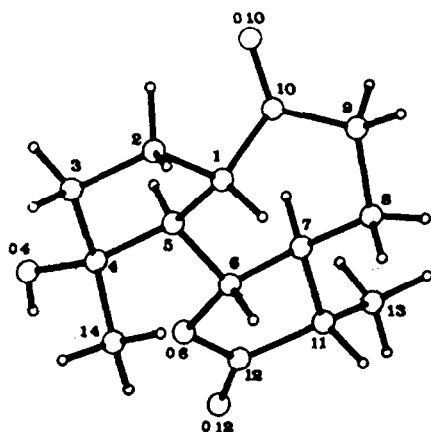


Figure 3. A view of compound 25 showing the atomic numbering.

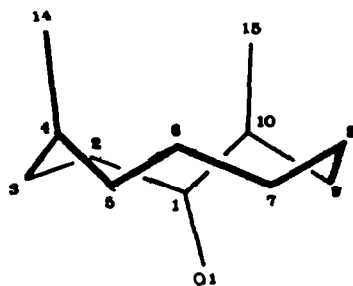
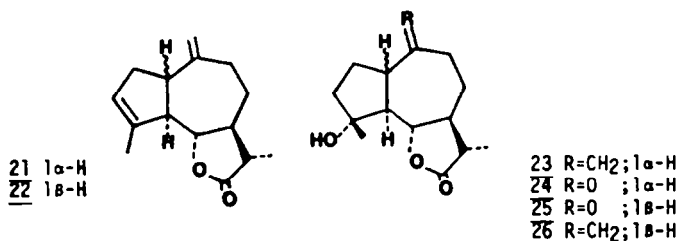


Figure 4. A projection of the ten-membered ring of compound 13 onto the plane normal to the best ring plane.

Table 1. Crystal Analysis Parameters at Room temperature

	Compound 13			Compound 23		
Formula	C ₁₅ H ₂₀ O ₃			C ₁₄ H ₂₀ O ₄		
Symmetry	Monoclinic, P2 ₁			Orthorhombic, P2 ₁ 2 ₁ 2 ₁		
Unit Cell Determination by l.s. Fit to:	45 reflections ($\theta < 45^\circ$)			38 reflections ($\theta < 40^\circ$)		
Unit Cell Dimensions	11.639(1)	12.274(1)	10.593(1)	22.52(1)	10.386(2)	5.689
	90.0	113.040(4)	90.0	90.0	90.0	90.0
Packing: V(A ³), Z	1392.6(1) 4			1330.6(7) 4		
Dc(g.cm ⁻³), M, F(1000)	1.184	248.32	536	1.260	252.31	544
Technique	Four-circle diffractometer, PH1100, bisecting geometry, CuK α , graphite oriented monochromator, /2 θ scans, scan width 1.5 $^\circ$, detector apertures 1" x 1", up θ max. 60 $^\circ$, 1 min./reflex.					
Number of Reflections (Friedel pairs)						
Independent	2162			1129		
Observed	1462 [2 σ (I) criterion]			1001 [2 σ (I) criterion]		
Standard Reflections	2 reflections every 90 minutes, no variation					
Solution and Refinement	MULTAN 80 [21], XRAY-76 [22], l.s. on F's					
H Atoms	Difference synthesis					
w-Scheme	Empirical, giving no trends in $\langle w 2F \rangle$ vs. $\langle Fo \rangle$ and $\langle \sin \theta / \lambda \rangle$					
Final R and Rw	0.073	0.098		0.066	0.075	
Scattering Factors	Int. Tables for X-Ray Crystallography [23]					
Anomalous Dispersion	Int. Tables for X-Ray Crystallography [23]					

1-epigallocatechin 11 suffered a biomimetic-type cyclization to the trans-guaianolide 22 (18%) when treated with TsCl in pyridine. The structure and stereochemistry of 22 were established by comparison with 21; the most important differences between the ¹H-NMR spectra of 21 and 22 are those of the H-14 signals (two broad singlets in 22; one broad singlet in 21). The structure of 22 was definitively established by comparison with an authentic sample prepared from 23¹⁴.



The ozonolysis of 23 gave ketone 24 (41%) which was converted into its epimer 25 by treatment with a 0.1 N solution of sodium hydroxide in methanol giving a 7:3 mixture of the ketones 25:24 which could be separated by fractionated crystallization. An X-ray diffraction study of 25 (Figure 3) showed the *trans* union of rings A and B. A list of bond lengths, bond angles and torsional angles for compound 25 is given in Table 3. The seven-membered ring presents a somewhat distorted twisted-chair conformation of the type TC₁¹⁵ and can be described as having a pseudo-twofold axis passing through C(9) and the midpoint of the

Table 2. Selected bond lengths (Å), bond angles ($^{\circ}$) and torsion angles ($^{\circ}$) for compound 13

		n=1	n=2			n=1	n=2
C(x1)-C(x2)		1.52(1)	1.54(1)	C(x7)-C(x8)		1.53(1)	1.50(1)
C(x1)-C(x10)		1.49(1)	1.51(1)	C(x7)-C(x11)		1.52(1)	1.57(1)
C(x1)-O(x1)		1.20(1)	1.23(1)	C(x8)-C(x9)		1.55(1)	1.58(1)
C(x2)-C(x3)		1.56(1)	1.56(1)	C(x9)-C(x10)		1.51(1)	1.47(1)
C(x3)-C(x4)		1.50(1)	1.52(1)	C(x10)-C(x15)		1.32(1)	1.36(1)
C(x4)-C(x5)		1.34(1)	1.29(1)	C(x11)-C(x12)		1.51(1)	1.46(1)
C(x4)-C(x14)		1.49(1)	1.50(1)	C(x11)-C(x13)		1.52(1)	1.48(1)
C(x5)-C(x6)		1.49(1)	1.52(1)	C(x12)-O(x6)		1.37(1)	1.37(1)
C(x6)-C(x7)		1.53(1)	1.52(1)	C(x12)-O(x12)		1.20(1)	1.20(1)
C(x6)-O(x6)		1.47(1)	1.46(1)				
		n=1	n=2			n=1	n=2
C(x10)-C(x1)-O(x1)		119.7(7)	118.2(8)	C(x6)-C(x7)-C(x11)		112.9(5)	112.1(6)
C(x2)-C(x1)-O(x1)		119.4(7)	121.1(9)	C(x7)-C(x8)-C(x9)		114.2(6)	114.0(6)
C(x2)-C(x1)-C(x10)		120.9(6)	120.6(8)	C(x8)-C(x9)-C(x10)		116.3(6)	117.4(7)
C(x1)-C(x2)-C(x3)		108.7(8)	107.4(7)	C(x1)-C(x10)-C(x9)		116.5(6)	117.7(7)
C(x2)-C(x3)-C(x4)		109.7(8)	110.1(8)	C(x9)-C(x10)-C(x15)		121.3(7)	123.6(8)
C(x3)-C(x4)-C(x14)		116.2(8)	116.0(7)	C(x1)-C(x10)-C(x15)		121.9(7)	118.3(7)
C(x3)-C(x4)-C(x5)		119.3(8)	118.9(9)	C(x7)-C(x11)-C(x13)		117.3(6)	116.0(6)
C(x5)-C(x4)-C(x14)		124.2(8)	124.7(8)	C(x7)-C(x11)-C(x12)		103.1(5)	104.6(6)
C(x4)-C(x5)-C(x6)		125.3(6)	125.6(8)	C(x12)-C(x11)-C(x13)		113.4(6)	113.3(6)
C(x5)-C(x6)-O(x6)		110.8(5)	109.9(5)	C(x11)-C(x12)-O(x12)		130.4(7)	129.7(8)
C(x5)-C(x6)-C(x7)		114.8(6)	112.6(6)	C(x11)-C(x12)-O(x6)		109.9(6)	110.7(7)
C(x7)-C(x6)-O(x6)		103.5(5)	106.1(5)	O(x6)-C(x12)-O(x12)		119.8(7)	119.6(8)
C(x6)-C(x7)-C(x11)		101.2(5)	101.0(5)	C(x6)-O(x6)-C(x12)		108.8(5)	109.5(6)
C(x6)-C(x7)-C(x8)		116.5(6)	118.3(6)				
		n=1	n=2			n=1	n=2
C(x2)-C(x1)-C(x10)-C(x15)		-33(1)	-28(1)	O(x6)-C(x6)-C(x7)-C(x11)		-36(1)	-28(1)
O(x1)-C(x1)-C(x10)-C(x9)		-23(1)	-16(1)	O(x6)-C(x6)-C(x7)-C(x8)		-159(1)	-151(1)
C(x2)-C(x1)-C(x10)-C(x9)		153(1)	169(1)	C(x6)-C(x7)-C(x11)-C(x12)		32(1)	24(1)
C(x10)-C(x1)-C(x2)-C(x3)		-88(1)	-93(1)	C(x6)-C(x7)-C(x11)-C(x13)		158(1)	150(1)
O(x1)-C(x1)-C(x2)-C(x3)		89(1)	82(1)	C(x6)-C(x7)-C(x8)-C(x9)		-77(1)	-81(1)
O(x1)-C(x1)-C(x10)-C(x15)		150(1)	157(1)	C(x6)-C(x7)-C(x11)-C(x12)		158(1)	151(1)
C(x1)-C(x2)-C(x3)-C(x4)		58(1)	56(1)	C(x6)-C(x7)-C(x11)-C(x13)		-77(1)	-83(1)
C(x2)-C(x3)-C(x4)-C(x5)		-96(1)	-94(1)	C(x11)-C(x12)-C(x6)-C(x9)		167(1)	163(1)
C(x2)-C(x3)-C(x4)-C(x14)		77(1)	79(1)	C(x7)-C(x8)-C(x9)-C(x10)		84(1)	75(1)
C(x3)-C(x4)-C(x5)-C(x6)		162(1)	159(1)	C(x8)-C(x9)-C(x10)-C(x11)		-133(1)	-123(1)
C(x14)-C(x4)-C(x5)-C(x6)		-11(1)	-14(1)	C(x8)-C(x9)-C(x10)-C(x15)		54(1)	64(1)
C(x4)-C(x5)-C(x6)-C(x7)		-131(1)	-131(1)	C(x7)-C(x11)-C(x12)-O(x6)		-18(1)	-12(1)
C(x4)-C(x5)-C(x6)-O(x6)		113(1)	111(1)	C(x7)-C(x11)-C(x12)-O(x12)		110(1)	106(1)
C(x5)-C(x6)-O(x6)-C(x12)		150(1)	144(1)	C(x13)-C(x11)-C(x12)-O(x12)		32(1)	39(1)
C(x5)-C(x6)-C(x7)-C(x8)		80(1)	89(1)	C(x13)-C(x11)-C(x12)-O(x6)		-145(1)	-140(1)
C(x5)-C(x6)-C(x7)-C(x11)		-157(1)	-148(1)	C(x11)-C(x12)-O(x6)-C(x6)		-6(1)	-6(1)
C(x7)-C(x6)-O(x6)-C(x12)		27(1)	22(1)	O(x12)-C(x12)-O(x6)-C(x6)		176(1)	175(1)

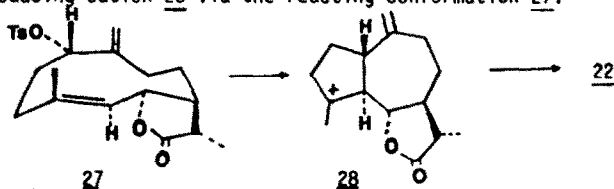
Table 3. Selected bond lengths (Å), bond angles (°) and torsion angles (°) for compound 25

C(1)-C(2)	1.552(8)	C(4)-C(5)	1.555(7)	C(6)-O(6)	1.477(6)	C(10)-O(10)	1.202(7)
C(1)-C(5)	1.567(7)	C(4)-C(14)	1.519(8)	C(7)-C(8)	1.540(8)	C(11)-C(12)	1.539(8)
C(1)-C(10)	1.505(8)	C(4)-O(4)	1.437(6)	C(7)-C(11)	1.537(7)	C(11)-C(13)	1.540(8)
C(2)-C(3)	1.538(11)	C(5)-C(6)	1.513(7)	C(8)-C(9)	1.541(8)	C(12)-O(6)	1.374(6)
C(3)-C(4)	1.538(9)	C(6)-C(7)	1.522(7)	C(9)-C(10)	1.525(9)	C(12)-O(12)	1.200(7)
C(5)-C(1)-C(10)	109.9(4)	C(1)-C(5)-C(4)	103.7(4)	C(1)-C(10)-C(9)	119.1(5)		
C(2)-C(1)-C(10)	113.8(5)	C(4)-C(5)-C(6)	116.6(4)	C(9)-C(10)-O(10)	118.9(5)		
C(2)-C(1)-C(5)	104.4(5)	C(1)-C(5)-C(6)	108.8(4)	C(1)-C(10)-O(10)	122.0(6)		
C(1)-C(2)-C(3)	106.8(5)	C(5)-C(6)-O(6)	110.0(4)	C(7)-C(11)-C(13)	116.1(4)		
C(2)-C(3)-C(4)	106.0(5)	C(5)-C(6)-C(7)	118.2(4)	C(7)-C(11)-C(12)	101.2(4)		
C(3)-C(4)-O(4)	107.0(4)	C(7)-C(6)-O(6)	102.8(4)	C(12)-C(11)-C(13)	112.5(5)		
C(3)-C(4)-C(14)	113.4(5)	C(6)-C(7)-C(11)	99.8(4)	C(11)-C(12)-O(12)	129.3(5)		
C(3)-C(4)-C(5)	101.6(4)	C(6)-C(7)-C(8)	112.5(4)	C(11)-C(12)-O(6)	110.0(4)		
C(14)-C(4)-O(4)	110.5(5)	C(8)-C(7)-C(11)	114.0(5)	O(6)-C(12)-O(12)	120.7(5)		
C(5)-C(4)-O(4)	110.8(4)	C(7)-C(8)-C(9)	113.4(5)	C(6)-O(6)-C(12)	107.6(4)		
C(5)-C(4)-C(14)	113.1(4)	C(8)-C(9)-C(10)	118.1(5)				
C(5)-C(1)-C(10)-O(10)	98.0(7)	C(1)-C(5)-C(6)-O(6)	175.6(4)				
C(2)-C(1)-C(10)-O(10)	-18.8(8)	C(5)-C(6)-O(6)-C(12)	154.8(4)				
C(5)-C(1)-C(10)-C(9)	-83.3(6)	C(6)-C(8)-C(7)-C(8)	76.7(5)				
C(2)-C(1)-C(10)-C(9)	160.0(6)	C(5)-C(6)-C(7)-C(11)	-162.1(4)				
C(2)-C(1)-C(5)-C(6)	-155.7(5)	C(7)-C(6)-O(6)-C(12)	30.4(5)				
C(10)-C(1)-C(5)-C(4)	-153.4(4)	O(6)-C(6)-C(7)-C(11)	-41.8(4)				
C(2)-C(1)-C(5)-C(4)	-31.0(5)	O(6)-C(6)-C(7)-C(8)	-163.1(4)				
C(5)-C(1)-C(2)-C(3)	8.2(7)	C(6)-C(7)-C(11)-C(12)	37.7(5)				
C(10)-C(1)-C(2)-C(3)	128.0(6)	C(6)-C(7)-C(11)-C(13)	159.8(5)				
C(10)-C(1)-C(5)-C(6)	81.9(5)	C(6)-C(7)-C(8)-C(9)	-83.3(6)				
C(1)-C(2)-C(3)-C(4)	18.0(7)	C(8)-C(7)-C(11)-C(12)	157.9(4)				
C(2)-C(3)-C(4)-C(5)	-36.9(6)	C(8)-C(7)-C(11)-C(13)	-80.0(6)				
C(2)-C(3)-C(4)-C(14)	84.8(6)	C(11)-C(7)-C(8)-C(9)	164.0(5)				
C(2)-C(3)-C(4)-O(4)	-153.1(5)	C(7)-C(8)-C(9)-C(10)	46.8(7)				
C(3)-C(4)-C(5)-C(1)	41.7(5)	C(8)-C(9)-C(10)-C(1)	28.0(8)				
C(3)-C(4)-C(5)-C(6)	161.0(5)	C(8)-C(9)-C(10)-O(10)	-155.2(6)				
C(14)-C(4)-C(5)-C(1)	-80.2(5)	C(7)-C(11)-C(12)-O(6)	-21.2(5)				
O(4)-C(4)-C(5)-C(1)	155.1(4)	C(7)-C(11)-C(12)-O(12)	159.1(6)				
C(14)-C(4)-C(5)-C(6)	39.2(6)	C(13)-C(11)-C(12)-O(12)	34.6(8)				
O(4)-C(4)-C(5)-C(6)	-65.6(5)	C(13)-C(11)-C(12)-O(6)	-145.8(5)				
C(4)-C(5)-C(6)-C(7)	175.3(4)	C(11)-C(12)-O(6)-C(6)	-5.5(5)				
C(1)-C(5)-C(6)-C(7)	-68.1(5)	O(12)-C(12)-O(6)-C(6)	174.2(5)				
C(4)-C(5)-C(6)-O(6)	59.0(5)						

C(5)-C(6) bond. Both five-membered rings have envelope conformation with C(4) [0.624(5) Å out of the plane] and C(7) [0.651(5) Å at the flap].

Treatment of 25 with sodium *t*-amlyoxide and triphenylmethylphosphonium bromide in toluene gave an 8:2 mixture of the alkenes 23 and 26, which, dehydrated with SO₂Cl-pyridine, generated the corresponding dienes 21 and 22 which were identified by comparison with authentic samples.

The transformation of 11 to 22 is a 5-endo-tet process and stereospecific¹⁶ which, taken together with the impossibility of isolating the intermediate sulphonic ester 14, strongly suggests that the cyclization takes place via a concerted process with the assistance of the double bond 4(5) producing cation 28 via the reacting conformation 27.

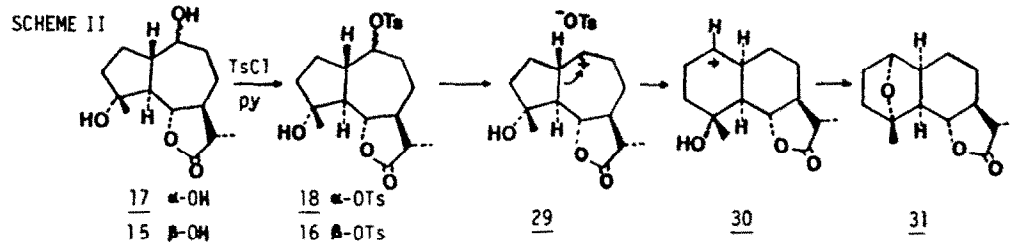


To the best of our knowledge this is the first time that a 1 α -hydroxy-*trans*-10(14)-germacradien-6,12-olide has been cyclized to form a *trans*-guaianolide, most probably via a reacting conformation similar to that adopted by the melampolides. It is of interest that the stereochemistry of the cyclization product is the same as shown by the few known natural *trans*-guaianolides, namely pleniradin¹⁷, galliardin¹⁸, neogalliardin¹⁸, florilenolin and its dihydroderivative¹⁹.

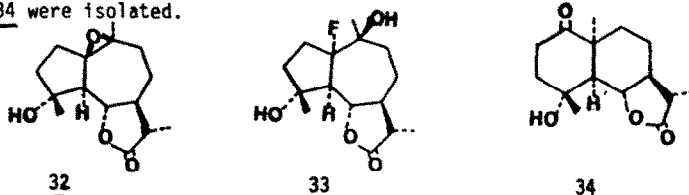
These findings strongly suggest that the biosynthesis of the *trans*-guaianolides may proceed via the melampolide pathway suggested by Parker *et al*².

In order to study the feasibility of a rearrangement similar to that formulated in Scheme I (6 from 5) we carried out the preparation of 15 and 17 by NaBH₄-EtOH reduction of 25, obtaining a 6:4 mixture. When MeOH-H₂O (8:2 v/v) was used as solvent, stereoselectivity was markedly increased and 17 (82%) was obtained with only traces of 15.

The tosylation of 17 led to epoxy-eudesmanolide 31 (25%) although the intermediate tosylate 18 could not be isolated. Identical results were obtained when the epimer 15 was treated in the same way. These data can be interpreted as shown in Scheme II.



Our results are completely in agreement with those obtained by Fischer *et al*²⁰ when the epoxyderivative 32 was treated with BF₃·Et₂O and the fluoride derivative 33 and the *cis*-eudesmanolide 34 were isolated.



It would seem that the σ bond (1,10) migrates more easily than H-1, which contradicts the theorized biogenetic hypotheses.

EXPERIMENTAL

Mp's were determined using a Kofler hot-plate apparatus and are uncorrected. I.r. spectra were recorded on a Perkin-Elmer 257 and NMR spectra were taken at 90 MHz on a Perkin-Elmer R-123 and at 200 MHz on a Bruker 200 SY, with CDCl_3 as solvent. Mass spectra were taken on a VG-Micromass ZAB-2F. Unless otherwise stated, column chromatography was carried out using Merck silica gel (0.065-0.2 mm).

Cyclization of 1-epigallocatechin 11

TsCl (250 mg) was added to a solution of 1-epigallocatechin 11 (314 mg) in minimal dry pyridine. After 24 h at -10°C , it was poured onto ice and water, frozen, washed with aqueous NaHCO_3 and NaCl -saturated solutions, extracted with CHCl_3 , dried, concentrated and chromatographed with hexane-EtOAc 9:1 as eluent, 22 (54 mg, 18.5%) was obtained as an oil: $^1\text{H-NMR}$: δ 5.45 (1H, bs, H-3); 5.05, 4.95 (1H each, s, H-14), 3.75 (1H, dd, $J=9$ and 10 Hz, H-6), 1.81 (3H, s, H-15); 1.20 (3H, d, $J=7$ Hz, H-13); i.r.: $\nu_{\text{max}}(\text{CHCl}_3)\text{cm}^{-1}$, 1770 (γ -lactone), 1600 (double bonds); h.r.m.s., m/z 232.1428 ($\text{C}_{15}\text{H}_{20}\text{O}_2$).

Ozonolysis of 23

A current of ozone (200 mg/h) was passed through a solution of 23, prepared according to González *et al.*¹⁴, (900 mg) in CH_2Cl_2 for 40 minutes at -78°C . Me_2S (1 ml) was added, the mixture was allowed to reach room temperature (4 h), and ice and a NaCl -saturated solution were added. It was extracted with CH_2Cl_2 , dried, concentrated and chromatographed with hexane-EtOAc 9:1 as eluent. 24 (376 mg, 41.5%) was obtained: mp 146-148°C; $[\alpha]_D^{25}$ -27.4° (CHCl_3 , 0.21); $^1\text{H-NMR}$: δ 3.86 (1H, dd, $J=9$ and 10 Hz, H-6); 1.29 (3H, s, H-15); 1.28 (3H, d, $J=7$ Hz, H-13); $^{13}\text{C-NMR}$: 56.38 (C-1), 21.10^a (C-2), 40.94 (C-3), 80.71 (C-4), 51.90^b (C-5), 85.27 (C-6), 50.50^b (C-7), 22.34^a (C-8), 42.84 (C-9), 209.50 (C-10), 39.18 (C-11), 177.53 (C-12), 12.49 (C-13), 23.00 (C-15). The signals marked ^a are interchangeable as are ^b. i.r.: $\nu_{\text{max}}(\text{CHCl}_3)\text{cm}^{-1}$, 3580 (OH), 1775 (γ -lactone), 1705 (carbonyl); m.s., m/z 252; 237; 234.

Epimerization of 24

A methanolic solution (0.1 N) of NaOH (5 ml) was added to a solution of 24 (54 mg) in MeOH (2 ml) and stirred for 30 minutes at 0°C . It was neutralized with an aqueous solution of HCl (5%), extracted with CH_2Cl_2 , dried and concentrated, yielding a 7:3 mixture of 25 and 24. Compound 25 was obtained by fractionated crystallization with hexane-EtOAc: mp 203-205°C; $[\alpha]_D^{25}$ 44.1° (CHCl_3 , 0.12); $^1\text{H-NMR}$: δ 4.08 (1H, dd, $J=9$ and 10 Hz, H-6); 1.38 (3H, s, H-15), 1.27 (3H, d, $J=7$ Hz, H-13); $^{13}\text{C-NMR}$: 53.78 (C-1), 28.67 (C-2), 40.31 (C-3), 79.88 (C-4), 50.54^a (C-5), 83.46 (C-6), 50.43^a (C-7), 23.00 (C-8), 44.30 (C-9), 210.58 (C-10), 41.06 (C-11), 177.35 (C-12), 13.47 (C-13). The signals marked ^a are interchangeable. i.r.: $\nu_{\text{max}}(\text{CHCl}_3)\text{cm}^{-1}$, 3580 (γ -lactone), 1705 (carbonyl); h.r.m.s., m/z 252.1508 ($\text{C}_{14}\text{H}_{20}\text{O}_4$).

X-ray Measurements

The experimental conditions and details of the X-ray analyses are given in Table 1. The final atomic parameters are given in Tables 4 and 5. In compound 13, both the atomic coordinates and the thermal parameters for the H-atoms were kept fixed during the last stages of refinement but in compound 25, only the thermal parameters. Both $hk1$ and $-h-k-1$ reflexions were used in the refinement. The absolute configuration²⁴ was confirmed using the 45 more relevant Friedel pairs with $F_c > 0.08$ and $F_o > 10\sigma(F_o)$ for compound 13 and with the 100 more relevant Friedel pairs with $F_c > 0.06$ and $F_o > 10\sigma(F_o)$ for compound 25. The averaged Bijvoet difference for 13 was 1074 for the right enantiomer as against 1226 for the wrong model, the corresponding averaged Bijvoet ratio being 1045 compared with 1063. For compound 25, the averaged Bijvoet difference was 0.446 for the right enantiomer as opposed to 0.521 for the wrong model, the corresponding averaged Bijvoet ratio being 1052 compared with 1069²⁵.

A view of the molecules with their atomic numbering is given in Figures 2 and 3.

Preparation of 22

A solution of freshly prepared sodium *t*-amyloxyde (0.5 M) in toluene was added to triphenylmethylphosphonium bromide (370 mg) and stirred under inert atmosphere until a yellow precipitate was formed. 25 (210 mg) dissolved in dry toluene (3 ml) was added and refluxed for 36 h then neutralized with aqueous solution of HCl (10%). After extraction with CH_2Cl_2 , it was dried, concentrated and chromatographed, using hexane-EtOAc 7:3 as eluent. A mixture of 23 and 26 (8:2) which could not be separated was obtained. This mixture was dissolved in dry pyridine (0.5 ml) and freshly distilled SOCl_2 (0.5 ml) was added, stirred for 5 minutes at 0°C , then poured over ice and water. An aqueous NaHCO_3 -saturated solution was added, the mixture was extracted with CH_2Cl_2 , dried, concentrated and chromatographed with hexane-EtOAc 9:1 as eluent. 21 and 22 were obtained in a 8:2 mixture which could not be separated and were identified by comparison with authentic samples using gas chromatography.

Reduction of 25

NaBH_4 (5 mg) was added to a solution of 25 (100 mg) in EtOH (5 ml), stirred for 30 minutes at 0°C , poured over ice and water, neutralized with aqueous solution of HCl (10%), concentrated at reduced pressure, extracted with Et₂O, dried, concentrated and chromatographed with hexane-EtOAc 4:6 as eluent. Compounds 17 (26 mg) and 15 (15 mg) were obtained. Compound 17: mp 158-160°C; $[\alpha]_D^{25}$ 19.8° (CHCl_3 , 0.21); $^1\text{H-NMR}$: δ 4.04 (1H, t, $J=4$ Hz, H-10); 3.90 (1H, dd, $J=9.5$ and 10 Hz, H-6); 1.31 (3H, s, H-15); 1.18 (3H, d, $J=6.5$ Hz, H-13); i.r.: $\nu_{\text{max}}(\text{CHCl}_3)\text{cm}^{-1}$, 3565 (OH), 1760 (γ -lactone); h.r.m.s., m/z 239.1283 ($\text{C}_{13}\text{H}_{19}\text{O}_4$, M+CH₃); 236.1414 ($\text{C}_{14}\text{H}_{20}\text{O}_3$, M+H₂O). Compound 15, oil: $^1\text{H-NMR}$: δ 4.58 (1H, dd, $J=9$ and 10 Hz, H-6); 3.90 (1H, d, $J=7$ Hz, H-10); 1.35 (3H, s, H-15); 1.19 (3H, d, $J=7$ Hz, H-13); i.r.: $\nu_{\text{max}}(\text{CHCl}_3)\text{cm}^{-1}$, 3580 (OH), 1760 (γ -lactone); h.r.m.s., m/z 254.1508 ($\text{C}_{14}\text{H}_{20}\text{O}_4$); 239.2590 ($\text{C}_{13}\text{H}_{19}\text{O}_4$); 236.1408 ($\text{C}_{14}\text{H}_{20}\text{O}_3$).

If MeOH-H₂O 4:1 was used instead of EtOH, as solvent, only 17 (82%) was obtained.

Table 4. Final atomic parameters for compound 13.

Thermal parameters as:

$$U_{eq} = (1/3) \cdot \sum [U_{ij} \cdot a_i^* \cdot a_j^* \cdot a_i \cdot a_j \cdot \cos(a_i, a_j)] \cdot 10^3$$

Atom	x/a	y/b	z/c	Ueq
C(11)	0.2359(6)	0.2285(6)	0.4543(7)	65(3)
C(12)	0.2571(6)	0.2438(6)	0.3226(8)	72(3)
C(13)	0.2773(7)	0.3675(7)	0.3045(7)	74(3)
C(14)	0.3862(6)	0.4085(5)	0.4268(6)	52(2)
C(15)	0.3655(6)	0.4575(5)	0.5285(6)	52(3)
C(16)	0.4607(6)	0.4750(5)	0.6697(6)	52(3)
C(17)	0.4200(6)	0.4398(5)	0.7849(6)	58(3)
C(18)	0.4292(6)	0.3180(6)	0.8186(7)	63(3)
C(19)	0.3234(6)	0.2481(6)	0.7139(7)	66(3)
C(110)	0.3428(6)	0.2124(5)	0.5877(8)	64(3)
C(111)	0.5056(6)	0.5089(6)	0.9036(6)	60(3)
C(112)	0.5219(7)	0.6111(7)	0.8337(7)	68(3)
C(113)	0.4641(8)	0.5306(8)	1.0205(7)	83(4)
C(114)	0.5128(6)	0.3802(7)	0.4318(7)	68(3)
C(115)	0.4450(7)	0.1601(6)	0.5969(8)	72(3)
O(11)	0.1315(4)	0.2347(7)	0.4511(6)	105(3)
O(16)	0.4881(4)	0.5919(4)	0.6964(4)	68(2)
O(112)	0.5546(6)	0.7004(5)	0.8789(6)	106(3)
C(21)	0.8756(9)	0.3487(8)	0.8811(8)	91(4)
C(22)	0.9201(8)	0.2522(10)	0.9809(8)	98(4)
C(23)	0.8742(9)	0.1457(9)	0.8952(10)	103(5)
C(24)	0.9243(8)	0.1402(7)	0.7831(8)	77(3)
C(25)	0.8560(7)	0.1763(6)	0.6626(7)	68(3)
C(26)	0.9033(6)	0.2113(6)	0.5539(6)	58(3)
C(27)	0.8524(5)	0.3214(5)	0.4919(6)	54(2)
C(28)	0.9237(7)	0.4215(6)	0.5592(7)	72(3)
C(29)	0.8997(8)	0.4600(7)	0.6895(8)	82(4)
C(210)	0.9591(7)	0.3967(6)	0.8169(7)	68(3)
C(211)	0.8457(6)	0.3066(6)	0.3423(7)	61(3)
C(212)	0.8364(6)	0.1886(8)	0.3196(8)	72(3)
C(213)	0.7461(7)	0.3687(8)	0.2339(7)	85(3)
C(214)	1.0571(10)	0.1039(9)	0.8267(10)	113(5)
C(215)	1.0844(8)	0.3890(9)	0.8864(9)	101(4)
O(21)	0.7689(7)	0.3841(8)	0.8447(7)	129(4)
O(26)	0.8623(4)	0.1340(4)	0.4406(5)	70(2)
O(212)	0.8085(5)	0.1379(6)	0.2144(6)	95(3)

Table 5. Final atomic parameters for compound 25.

Thermal parameters as:

$$U_{eq} = (1/3) \cdot \sum [U_{ij} \cdot a_i^* \cdot a_j^* \cdot a_i \cdot a_j \cdot \cos(a_i, a_j)] \cdot 10^3$$

Atom	x/a	y/b	z/c	Ueq
C(1)	0.0819(2)	0.6772(5)	0.4734(10)	43(2)
C(2)	0.0766(4)	0.5400(7)	0.3675(14)	70(3)
C(3)	0.1183(4)	0.5352(7)	0.1540(12)	65(2)
C(4)	0.1616(2)	0.6487(5)	0.1837(9)	42(2)
C(5)	0.1203(2)	0.7534(5)	0.2907(8)	32(1)
C(6)	0.1504(2)	0.8642(5)	0.4158(9)	33(1)
C(7)	0.1100(2)	0.9716(5)	0.5028(9)	37(2)
C(8)	0.0761(3)	0.9344(6)	0.7278(11)	50(2)
C(9)	0.0193(2)	0.8562(6)	0.6796(14)	59(2)
C(10)	0.0232(2)	0.7436(6)	0.5081(11)	52(2)
C(11)	0.1561(2)	1.0790(5)	0.5384(11)	40(2)
C(12)	0.1969(2)	1.0561(5)	0.3320(10)	45(2)
C(13)	0.1318(3)	1.2175(6)	0.5450(16)	60(2)
C(14)	0.2146(3)	0.6172(7)	0.3386(12)	56(2)
O(4)	0.1809(2)	0.6851(4)	-0.0476(6)	49(1)
O(6)	0.1907(1)	0.9320(3)	0.2525(7)	41(1)
O(10)	-0.0210(2)	0.7094(5)	0.4075(10)	91(2)
O(12)	0.2312(2)	1.1287(4)	0.2396(9)	65(2)

Tosylation of 17 and 15

To 17 (117 mg) dissolved in dry pyridine (1 ml) TsCl (105 mg) was added and the mixture kept at room temperature for three days, then poured over ice and water, washed with an NaHCO₃-saturated solution, extracted with CH₂Cl₂, dried, concentrated and chromatographed with hexane:EtOAc 7:3 as eluent. 32 (37 mg) and 18 (17 mg) were obtained. Compound 18 decomposed rapidly. Compound 32: mp 141-142°C, [α]_D²⁰ -64.1° (CHCl₃, 0.18), ¹H-NMR: δ 4.05-3.91 (2H, complex, H-1 and H-6), 1.35 (3H, s, H-15); 1.21 (3H, d, J=7 Hz, H-13), ¹H-NMR (C₆D₆): δ 3.40-3.25 (1H, complex, H-1); 3.20 (1H, dd, J=9 and 10 Hz, H-6); 1.11 (3H, s, H-15); 0.90 (3H, d, J=7 Hz, H-13); i.r.: ν_{max} (CHCl₃) cm⁻¹, 1765 (γ-lactone); h.r.m.s., m/z 236.1394 (C₁₄H₂₀O₃); 221.1216 (C₁₃H₁₇O₃); 218.1322 (C₁₃H₁₆O₂).

When 15 was tosylated (24 h) product 32 (35%) was also obtained.

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